

# THE PREVALENCE OF *MORAXELLA BOVIS* IN CLINICALLY NORMAL CATTLE EYES: ITS SIGNIFICANCE TO INFECTIOUS BOVINE KERATO-CONJUNCTIVITIS

<sup>1</sup>G.O. EGWU, <sup>2</sup>L.T. ZARIA, <sup>2</sup>N.L. IMAN

<sup>1</sup>Department of Vet. Medicine

<sup>2</sup>Department of Vet. Microbiology & Parasitology

Faculty of Veterinary Medicine,

University of Maiduguri

P.M.B. 1069, Maiduguri.

(Received 20 May 1991, accepted 6 August 1991).

## ABSTRACT

In a study to determine the prevalence of *Moraxella bovis* in clinically normal cattle eyes in Maiduguri area of Nigeria, 35 adult cattle (70 eyes) and 25 calves (50 eyes) were sampled. Three each of the adult and young cattle making a total of 6 (5%) were positive for *M. bovis* (the aetiological agent of infectious bovine Keratoconjunctivitis) out of the 120 eyes sampled. *Branhamella catarrhalis*, *Branhamella mucosa* and *Escherichia coli* were each isolated respectively, from 2 (1.6%) of the 120 samples. The other bacterial species isolated were *Bacillus* (5; 4.2%), *Corynebacterium* (17; 14.2%), *Streptococcus* (9; 7.7%) and *Staphylococcus* (21/17.5%), could be regarded as commensals. The epidemiological implications of finding *M. bovis* in clinically normal cattle eyes are discussed.

## INTRODUCTION

Infectious bovine kerato-conjunctivitis (IBKC) is recognised world-wide as a common condition affecting the eyes of cattle. Various other names have been given to the disease such as pink eye (Scott, 1957) and New forest disease (Bedford, 1976).

*Moraxella bovis* is considered the aetiological agent of infectious bovine keratoconjunctivitis (Adinarayanan and Singh, 1961; Pugh and Hughes, 1975). The disease is considered to be of major economic importance to the cattle industry in some parts of the world (Killinger *et al.*, 1977; Harris *et al.*, 1980).

Nigeria Journal of Animal Production 18 (1991)

In Nigeria, there appears to be little or no report on the isolation of *M. bovis* from clinically normal cattle eyes, although the prevalence of serum antibodies to *M. bovis* has been reported (Makinde *et al.*, 1985). Therefore, this paper reports on the isolation of *M. bovis* from normal cattle eyes and highlights on the significance of the findings.

## MATERIALS AND METHOD

### Sources of Samples

Samples were taken from each eye of thirty five (70) samples adult cattle from the Maiduguri cattle market, and twenty five (50 samples) calves in the University of Maiduguri Teaching and Research Farm. The breeds involved were white Fulani, Wadara, Kuri and their crosses.

### Collection of Samples

Only eyes which showed no clinical disease (unaffected; grade 0) were sampled.

The upper and lower fornices of the conjunctivae were sampled with a transwab<sup>(R)</sup> (Medical Wire Equipment, England) containing a sterile transport medium.

### Media Cultivation and Identification

After sampling, and on return to the laboratory each swab was immediately streaked onto blood agar plates containing 5% bovine blood. The plates were then incubated at 37°C aerobically for 24hrs, and a further 48hrs for those plates that showed scantily or no growth. Representative

colonies of both gram positive and negative bacteria on each plate were Gram stained, including those that showed typical morphological and cultural characteristics of *Moraxellae* or *Branhamellae* (*Neisseriae*) and were further subcultured for purity onto bovine blood agar and reincubated at 37°C for 24 hr. Other isolates which included *Bacillus*, *Corynebacterium*, *Streptococcus* and *Staphylococcus* species were identified to the genus level based on morphology, and routine biochemical and sugar reactions according to the methods of Cowan and Steel (1974).

Colonies culturally resembling *Moraxella* and *Brahamella* species were further examined fermentatively and also biochemically using catalase, oxidase, litmus milk and the ability to produce haemolysin as described by Mcfaddin (1980) and Bergey (1984). *Moraxella* and *Brahamella* species were then differentiated according to the methods of Fraser and Gilmour (1979).

## RESULTS

Table I shows the number of gram negative samples isolated from 120 clinically normal cattle eyes. Thirteen gram negative isolates were positive out of the 120 eye samples, of which *M. bovis* was isolated from 6 (5%) of the total eye samples. Of the positives samples for *M. bovis*, 3 (5%) were obtained from calves and the other 3 (5%) was obtained from the adult cattle. All the isolated *M. bovis* showed typical fermentative and biochemical characteristic of *Moraxella* as described by Fraser and Gilmour (1979). *Brahamella catarrhalis* was isolated from 2 (1.6%) of the 120 samples. Two isolates (1.6%) of *Branhamella mucosa* and 1 (0.8%) of *Branhamella pharyngis* were obtained from the 120 samples, whilst *Escherichia coli* was positive in 2 (1.6%) of the total samples.

The gram positive bacterial species also isolated from the 120 samples were *Bacillus* (5, 4.2%), *Corynebacterium* (17; 14.2%), *Streptococcus* (9; 7.5%) and *Staphylococcus* (21; 17.5%).

## DISCUSSION

This study has shown that a variety of micro-organisms exist in cattle eyes similar to the obser-

vations of Barber et al (1986). Since *M. bovis* is considered as the aetiological agent of IBKC (Adinarayananand Singh, 1961; Pugh and Hughes, 1975), the isolation of this organism from clinically normal cattle eyes in this study, indicate that these animals may have been recovered carriers following a previous infection. It is also likely that these positive animals may have been in their subclinical stage of the disease (Infectious bovine kerato-conjunctivitis).

Only few isolates (6) of *M. bovis* were recovered from the 120 sample, perhaps because these positive animals had just recently been infected with *M. bovis* prior to sampling, or that local ocular immune responses might have neutralised the organism in some infected eyes especially the secretory IgA(s), which are known to predominate in affected eyes following infection with *M. bovis* (Pederson and Nansen, 1972; Duncan et al, 1972). However, the presence of this organism in cattle eyes indicate that these animals can act as carriers and may constitute a source of infection to other animals in the flock or those newly introduced, which can lead to new outbreaks or recrudescence of the disease.

The other bacterial species isolated in this study were similar to those of spradbrow (1967) and Wilcox (1970). However, it is not known what role these other organisms play in the pathology of the disease, although they have not been reported to cause ocular disease in cattle (Bergey, 1984). It is also likely that these organisms may act as secondary invaders or commensals in normal or affected eyes.

This study has shown that *M. bovis* can occur subclinically, or in normal cattle eyes with some epidemiological implications. The work reported in this study can be regarded as preliminary, as further work is in progress to monitor infected eyes in some herds by repeated samplings, in order to ascertain the true prevalence of *M. bovis* in these eyes.

## REFERENCES

ADINARAYANAN, N. and SINGH, S.B.

TABLE 1: FERMENTATION AND BIOCHEMICAL CHARACTERISATION OF GRAM-NEGATIVE BACTERIA ISOLATED FROM 120 CLINICALLY NORMAL CATTLE EYES.

Number of Samples	Haem.	Growth on mac.	Oxidasc	Catalasc	Nitrate	Litmus milk	Liquefac tion of Grlatint	Glucose	Maltose	Lactose	Xylose	Identification
5	-	-	+	-	+	NP	-	-	-	-	-	<i>B. Catarhalis</i>
11	-	-	+	+	+	NP	-	-	-	+	-	<i>B. mucosa</i>
17	B	-	+	-	-	P	+	-	-	-	-	<i>B. bovis</i>
18	-	+	-	+	+	-	+	+	+	+	+	<i>E. coli</i>
22	-	-	+	+	-	NP	-	+	-	-	-	<i>B. pharyngis.</i>
35	-	+	-	+	+	NP	-	+	+	+	-	<i>E. coli</i>
43	B	-	+	+	-	P	+	-	-	-	-	<i>M. bovis</i>
44	B	-	+	+	-	P	+	-	-	-	-	<i>M. bovis</i>
50	B	-	+	+	-	P	+	-	-	-	-	<i>M. bovis</i>
72	-	-	+	+	-	P	+	-	-	-	-	<i>M. bovis</i>
36	-	-	+	+	+	AC	-	+	+	+	+	<i>B. mucosa</i>
106	+	-	+	+	+	AC	-	-	-	-	-	<i>B. catarhalis</i>
116	-	-	+	+	-	P	+	-	-	-	-	<i>M. bovis</i>

Haem = Haemolysis on bovine blood agar

B = Clear zone of haemolysis

Mac = MacConkey agar

+

= Positive

-

= Negative

P

= Peptonization

NP

= No peptonization

AC

= Acid.

- (1961). Infectious bovine keratitis with special reference to isolation of *Moraxella bovis*. *Veterinary Record*, 73: 694-696.
- BARBER, D.M.L., JONES, G.E. (1986). Microbial flora of the eyes of cattle. *Veterinary Record*, 118: 204-206.
- BEDFORD, P.G.C. (1976). Infectious bovine kerato-conjunctivitis: A review - *Veterinary Record*, 98: 134-135.
- BERGEY, D.H. (1984). *Bergey's Manual of Determinative Bacteriology*. Eds. H.A. Peter, N.S. Sneath, N.S. Mair, M.E. Sharpe and J.G. Holt, Vol. 2, pp. 189-190, Williams and Wilkins, Baltimore, U.S.A.
- COWAN, S.T. and STEEL, S.K. (1974). *Cowan and Steel's Manual for the identification of Medical Bacteria*, 2nd Edition. Cambridge University Press.
- DUNCAN J.R., WILKIE, B.N., HIEST F, and WINTER, H.J. (1972). The serum and secretory immunoglobulins of cattle: Characterisation and quantitation. *Journal of Immunology*, 108: 965-976.
- FRASER, J., and GILMUR, N.J.L. (1979). The identification of *Moraxella bovis* and *Neisseria ovis* from the eyes of cattle and sheep, *Research in Veterinary Science*, 27: 127-1128
- HARRIS, R.E., COOPER, B.S., STEFFERT, I. J. and BRICE, J.S. (1980). A survey of bovine infectious keratoconjunctivitis (pink eye) in beef cattle. *New Zealand-Veterinary Journal*, 28: 56-60.
- KILLINGER., A.H., VALENTINE, D. MANSFIELD, M.E., RICKETTS, G.E. CMARIK; G.F., NEUMANN, A.H. and NORTON, H.W. (1977). Economics impact of infectious bovine keratoconjunctivitis in beef calves, *Veterinary Medicine small animal Clinician*, 72: 618-620.
- MAKINDE, A.A., EZE, A.O., ONOVIRAN, O. and UMOR, I. (1985). Prevalence of serum antibodies to *Moraxella bovis* in cattle in Nigeria, *British Veterinary Journal*, 141: 643-646.
- MACFADDIN, J.F. (1980). *Biochemical Tests for the identification of Medical Bacteria* 2nd edition pp 64-115, Williams and Wilkins, Baltimore, U.S.A.
- PEDERSON, K.B. and NANSEN, P. (1972). Immunoglobulins in bovine lachrymal fluid. *Acta pathologica Microbiologica Scandinavica*. 80: 213-240.
- PUGH, G.W and HUGHENS, D.E. (1968). Experimental bovine infectious keratoconjunctivitis caused by sun lamp irradiation and *Moraxella bovis* infection: Correlation of haemolytic ability and pathogenicity. *American Journal of Veterinary Research*, 29: 835-839.
- SCOTT, G.C. (1957). The use of cortisone in the treatment of infectious keratoconjunctivitis (pink eyes) in cattle. *Journal of American Veterinary Medical Association*. 130: 257-259.
- SPRADBROW, P.B. (1967). A microbiological study of bovine conjunctivitis and keratoconjunctivitis. *Australian Veterinary Journal*. 43: 55-58.
- WILCOX, G.E. (1970). Bacterial flora of the bovine eye with special reference to the *Moraxell* and *Neisseria*. *Australian Veterinary Journal*, 46: 253-257.