

## Anaesthetic and electrocardiographic profiles in prolonged inhalation anaesthesia in Nigerian indigenous dogs

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### Abstract

*Electrocardiograph during anaesthesia provides information on cardiac electrical activity which may affect cardiac function. This study was carried out to evaluate the effect of prolonged volatile anaesthesia using halothane and isoflurane on the electrographic and anaesthetic profiles in Nigerian dogs. They were prepared for anaesthesia and connected to a 5-lead patient monitor (GMI®). Venous access was secured and lactated ringer's solution administered at a maintenance flow rate of 5mlkg<sup>-1</sup>. Tracheae were intubated following induction with thiopentone and anaesthesia maintained with 0.5% halothane (group A) and 2.0% isoflurane (group B) in 2liters/minute oxygen respectively with the animals breathing spontaneously. The readings were taken, prior to induction of anaesthesia (control) and every 30 minutes thereafter for six hours during anaesthesia. Data were presented as mean and standard deviation. Differences were considered significant at  $p < 0.05$ . The values of S-T segment (sec) and P-waves (sec) were both low,  $0.068 \pm 0.013$  and  $0.064 \pm 0.017$  and  $0.020 \pm 0.001$  and  $0.020 \pm 0.001$  respectively for isoflurane and halothane. However the values of P-waves (mV) and P-R intervals (sec) were significantly higher in group A, ( $0.193 \pm 0.136$  and  $0.046 \pm 0.01$ ) compared to group B,  $0.193 \pm 0.98$  and  $0.039 \pm 0.01$  respectively. Isoflurane produced prolongation of Q-T intervals (sec) and QRS complex (sec),  $0.123 \pm 0.018$  and  $0.023 \pm 0.008$  compared to halothane,  $0.121 \pm 0.023$  and  $0.021 \pm 0.005$ . The ECG parameters measured revealed no adverse effect of halothane or isoflurane on the heart in Nigerian local dogs. There was prolonged QT interval in group B. In conclusion, halothane appears a better drug of choice in prolonged anaesthesia in Nigerian dogs.*

**Keywords:** Anaesthetic, electrographic profiles, Nigerian indigenous dogs

### Introduction

Electrocardiogram (ECG) is a test that measures the electrical activity of the heart. The heart is a muscular organ that beats in rhythm to pump the blood through the body (Davey, 2011). The signals that make the heart's muscle fibres contract come from the sino-atrial node, which is the natural

pacemaker of the heart (Larsson, 2010). The ECGs from normal, healthy hearts have a characteristic shape. Any irregularity in the heart rhythm or damage to the heart muscle can change the electrical activity of the heart so that the shape of the ECG is changed. Monitoring the Electrocardiograph during anaesthesia provides information on cardiac

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electrical activity which may affect cardiac function. Nigerian local dogs are indigenous to Nigeria; they are mainly used for hunting in the wild by the local people and are also used as guard dogs and pets in the urban areas (Olayemi *et al.*, 2009). A number of scholars have worked on this breed of animal, reporting variations in clinical parameters between this breed and other breeds in the temperate (Awah and Nottidge, 1998, Olayemi *et al.*, 2009). Some have reported variations in the anatomical make-up of this breed and some other foreign breed (Nnaji *et al.*, 2010; Igado, 2011). This study evaluated the effects of halothane and isoflurane anaesthesia on anaesthetic indices and the ECG in Nigerian local dogs

### **Materials and methods**

The study was conducted at the Veterinary Teaching Hospital, College of Veterinary Medicine, Federal University of Agriculture, Abeokuta, Nigeria. Clinical trials were carried out on twenty healthy adult dogs, averaging  $11.05 \pm 1.88$  kg. They were grouped into two by simple randomization. Group A was treated with halothane and group B with isoflurane. Five self-adhesive electrodes were attached to the median sides of the skin of the fore limbs at the elbows, the hind limbs at the stifles and the thorax at the left sixth inter-coaster space near the sternum. These areas were clipped to prevent air trapping between the skin and electrodes, the electrodes were placed on the clipped areas and the 5 leads of the multi-parameter patient monitor (GMI<sup>®</sup>) were connected to the patient. Records of the electrocardiograph parameters, the heart and pulse rates were taken with speed 50mm/sec. Venous access was secured at the left cephalic vein and lactated ringers solution was administered at a maintenance

flow rate of  $5 \text{mlkg}^{-1}$  (AAHA/AAFP; Davis *et al.*, 2013). Premedication was achieved with intravenous injection of 2% Atropine Sulphate (Martindale Pharmaceutical, UK) at  $0.04 \text{mgkg}^{-1}$ , thirty minutes afterwards anaesthesia was induced with 2.5% thiopental sodium (May & Baker Ltd) at  $15 \text{mgkg}^{-1}$  intravenously. Tracheae were intubated with the aid of laryngoscope using appropriate sized endotracheal tubes (5.5-8.5ID) and anaesthesia was maintained with 0.5% halothane (Pharco Pharmaceutical, Egypt) (group A) and 2.0% isoflurane (Nicholas Pharmaceuticals India Ltd, India) (group B) in 2liters/minute oxygen and closed circle breathing system. The animals were placed in right lateral recumbency, breathing spontaneously. The readings were taken, one, prior to induction of anaesthesia (control) and every 30 minutes thereafter for six hours during anaesthesia on the patient monitor.

Anaesthetic indices taken in the course of this study included the time to intubation, time to extubation, time to sternal and time to stand. Monitoring was undertaken throughout anaesthetic period. Data obtained were analyzed using one-way ANOVA for repeated measures. Differences were considered significant at  $p = 0.05$ .

### **Result and discussion**

The quality of anaesthesia obtained in this study for both drugs, halothane and isoflurane were good. Higher dose of isoflurane was required to maintain surgical plane of anaesthesia compare to halothane in this breed 0.5% and 2.5% respectively. The dose of halothane used in this study agrees with the works of Allen *et al.*, 1998 and Muir *et al.*, 2007 but the dose of isoflurane used to maintain anaesthesia in this study disagrees with the work of these scholars.

Recovery from anaesthesia was

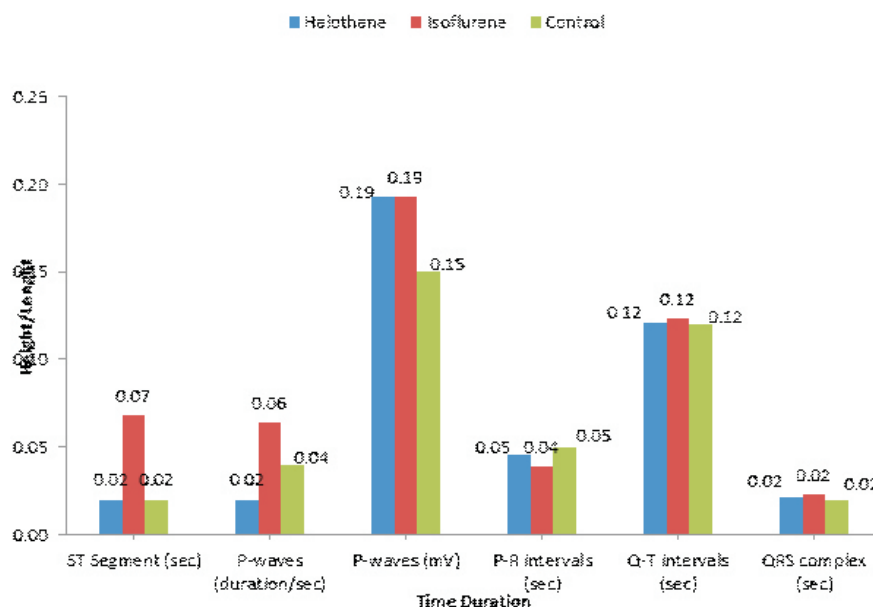
significantly ( $p > 0.05$ ) shorter in the halothane group,  $23.5 \pm 15.8$  minutes compare to the isoflurane group,  $34.2 \pm 27.8$ . This is believed to be due to the dose of isoflurane group used for maintenance of anaesthesia. Recovery was smooth and uneventful in both groups.

The side effects observed in this study were breath holding, shivering, oedematous head and tongue in a dog in the isoflurane group and hypothermia in one of the animals in the halothane group. There were no incidences of difficult intubation, kinked tube or obstruction due to excessive secretion, morbidity or mortality recorded in this study.

The study showed that all the ECG parameters measured were within normal range for the two drugs considered except for the values of S-T segment (sec) and P-waves (Duration)(sec) that were both low,  $0.068 \pm 0.013$  sec and  $0.064 \pm 0.017$  sec and  $0.020 \pm 0.001$  sec and  $0.020 \pm 0.001$  sec

respectively for isoflurane and halothane respectively as reported by Goodwin *et al.*, 1996. Although the value of P-R intervals (sec) was significantly higher in the halothane group,  $0.046 \pm 0.01$  sec compared to isoflurane group,  $0.039 \pm 0.01$  sec. both values are low, this could be due to transient myocardial ischaemia as seen in lowered value of the ST segment. (the lower the value compare to the baseline value, the higher the value of the PR interval).

However the value of P-waves (mV) was significantly higher in halothane group,  $0.193 \pm 0.136$  mV and  $0.046 \pm 0.01$  sec compared to isoflurane group,  $0.193 \pm 0.98$  mV and  $0.039 \pm 0.01$  sec respectively while the values of Q-T intervals (sec),  $0.123 \pm 0.018$  sec and QRS complex (sec),  $0.023 \pm 0.008$  sec were significantly prolonged in group B, compared to group A,  $0.121 \pm 0.023$  sec and  $0.021 \pm 0.005$  sec respectively, (Mitchalodis *et al.*, 1998; Niyazi *et al.* 2001; Brooker *et al.*, 2003).



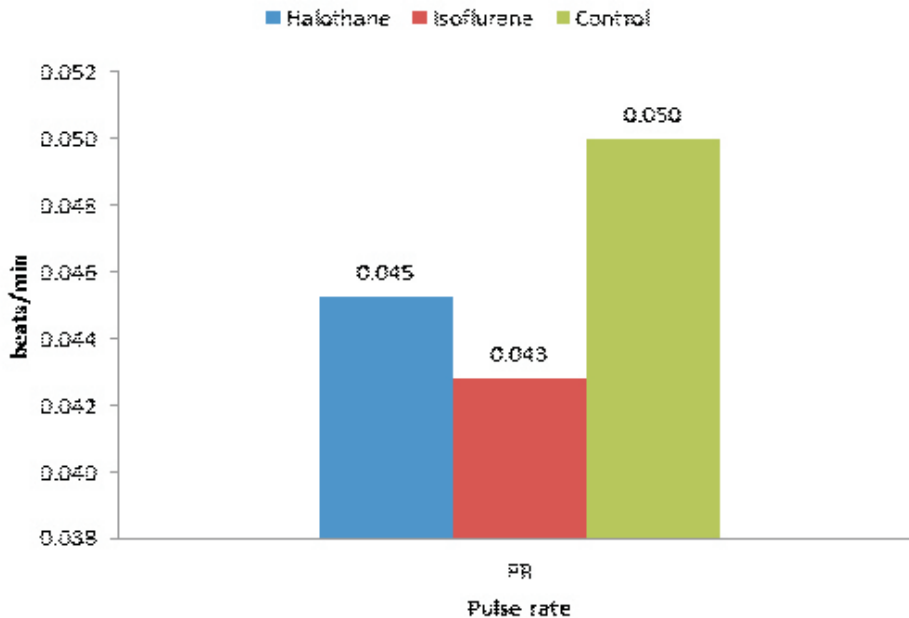
**Fig. 1: Effects of Halothane and Isoflurane Anaesthesia on the Electrocardiograph of the adult Nigerian dog**

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There was marked increase in the heart rates in both drugs studied over the six hours period of study,  $147 \pm 23.82$  beats/minute compare to the baseline value of  $91 \pm 30.25$  beats/minute in the halothane group and in isoflurane group,  $145 \pm 23.06$  beats/minute compare to baseline value, of  $107 \pm 16.62$  beats/minute.

Although there was later a decline to  $117.5$  beats/minute this was still significantly different from the control value. However, there was no significant difference between groups A and B. The findings agrees with the report of Hikasa *et al.*, 1998a and Hikasa *et al.*, 2010 in an experiment carried

out on goats, they reported that using 1-1.5 MAC of halothane, isoflurane and sevoflurane in goat gave an increase in heart rate in each group from baseline and the significant increase observed during anaesthesia is due to the influence of atropine sulphate premedication. The heart rates in both groups returned to normal values towards the last two hours of the experiment  $120.23 \pm 23.82$  beats/minutes in the halothane group and isoflurane group,  $120.75 \pm 23.06$  beats/ minutes compared. This is in agreement with the findings of Williams; Picker *et al.*, 2001.



**Fig. 2: Effects of Halothane and Isoflurane Anaesthesia on Pulse rate of the adult Nigerian dog**

#### Conclusion

The study on the electrocardiograph revealed no adverse effect of halothane or isoflurane on the heart in Nigerian indigenous dogs different from what is seen in literature in other breeds of dogs. However, the atrial depolarization, start of

atrial depolarization, ventricular depolarization and time taken by ventricular depolarization and repolarization were higher in isoflurane treated group than in halothane treated group. Judging by the responses however, halothane seemed to be a better inhalation

anaesthetic in a prolonged anaesthetic protocol in this breed of dogs (especially in patient with long QT syndrome). It is recommended that more work be done on this breed to develop a safer inhalation anaesthetic protocol considering that the dose of isoflurane used in this study to maintain anaesthesia is relatively high.

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#### **Reference**

- Allen, D. G., Pringle, J. K., Pasloske, K. and Smith, D. A. 1998.** Handbook of Veterinary Drugs. 2<sup>nd</sup> edn Lippincott-Raven, New York. pp. 825-826.
- Awah, J. N. and Nottidge, H. O. 1998.** Serum biochemical Parameters in Clinically Healthy Dogs in Ibadan. *Tropical Veterinarian* 16, pp.123-129.
- Booker, P. D., Whyte, S. D. and Ladusaus, E. J. 2003.** Long QT syndrome and anaesthesia. *British Journal of Anaesthesia*, 90(3), pp. 349-366.
- Davis, H., Jessen, T., Johnson, A., Knowles, P., Meyer, R., Rucinsky, R. and Shafford, H. 2013.** AAHA/AAFP Fluid therapy guidelines for dogs and cats. *Journal of American Animal Hospital Association*, 149(3), pp. 149-159.
- Goodwin, J. K., Atkins, C. and Tilley, L. 1996.** Electrocardiography. In: Diagnostic Cardiology. Practical Resource Manual, Companion Animal Cardiology Series I, Module Lifelearn<sup>®</sup> Inc. pp.1-12
- Hikasa, Y., Okuyama, K., Kakuta, T., Takase, K. and Ogasawara, S. 1998a.** Anaesthetic potency and cardiopulmonary effects of sevoflurane in goats: comparison with isoflurane and halothane. *Canadian Journal of Veterinarian Research*. 62, pp. 299–306.
- Hikasa, Y., Yamashita, M., Takase, K. and Ogasawara, S. 2010.** Prolonged sevoflurane, isoflurane and halothane anaesthesia in oxygen using rebreathing or non-rebreathing system in cats. *Journal of Veterinary Medicine Series A45*, pp.559-575.
- Igado, O. O. 2011.** Gross morphometric study of the eyeball and tongue of the Nigerian local. *Italian Journal of Anatomy and Embryology*, 116(1), pp.104-110.
- Larsson, H. P. 2010.** How is the heart rate regulated in the sinoatrial node?. Another piece to the puzzle. *Journal of General Physiology*, 136(3), pp.237-241. Doi: 10.1085/jcp.201010506
- Muir, W. W., Hubbell, J. A. E. and Bednarski, R. M. 2007.** Introduction to anaesthesia. In: Handbook of Veterinary Anesthesia 4<sup>th</sup> edn. St Louis, Mosby. pp1-10.
- Michaloudis, D., Fraidakis, O., Lefaki, T., Kanakoudis, F. and Askitopoulou, H. 1998.** Anaesthesia and the QT interval in humans: effects of halothane and isoflurane in premedicated children. *European Journal of Anaesthesiology*, 15 (6), pp. 623-628.
- Niyazi, G., Ismail, K., Cengiz, B. D., Mehmet, B. Beyhan, E. and Cevat, T. 2001.** The effects of volatile

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- anesthetics on the Q-Tc interval. *Journal of Cardiothoracic and Vascular anaesthesia*, 15(2), pp.188-191.
- Nnaji, T. O., Ibeagi I. O., Udegbunam, S. O. and Udegbunam, R. I. 2010.** Frontosagittal Index, Vertebral Index and Vertical Diameter of Thorax in Mongrel Dogs. *Animal Science Reporter*, 4(1), pp.3-8.
- Olayemi, F. O., Azeez, I. O., Ogunyemi, A. and Ighagbon, F. O. 2009.** Study on the erythrocyte values of the Nigerian indigenous dogs. *Folia Veterinarian*, 53, pp. 65-67.
- Picker, O., Scheeren, T. W. L. and Atnolt, J. O. 2001.** 'Inhalation anaesthetics increase heart rate by decreasing cardiac vagal activity in dogs'. *British Journal of Anaesthesia*, 87, pp. 748-754.
- William, C. 2011.** ECG Interpretations. *Animal Specialty and Emergency Center (ASEC)*. pp. 1-43. [www.asecvets.com/wp-content/uploads/2011/06/ECG\\_interpretionpdf](http://www.asecvets.com/wp-content/uploads/2011/06/ECG_interpretionpdf).

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