

Haematological Response and Carcass Yield of Broiler Chickens Administered Spotted Pumpkin (*Lagenaria Breviflora* Robert) Fruit Extracts

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Abstract

Over the years, therapeutic application of plants has been used in folkloric medicine to explore natural alternatives to antibiotic growth promoters due to the development of resistant bacteria strains and presence of drug residues in poultry meat. This study investigated the effects of concentrations and frequencies of administration of *Lagenaria breviflora* fruit extract (LBFE) on the haematological indices and carcass yield of broiler chickens. A total of 384 1-day-old male Arbor acres chicks were randomly distributed into six treatments; T1: control, T2: 75 g of LBFE/litre for 3days/week, T3: 75 g of LBFE/litre for 5days/week, T4: control, T5: 125 g of LBFE/litre for 3days/week and T6: 125 g of LBFE/litre for 5days/week. The experiment was laid out in a 2×3 factorial and data were subjected to Completely Randomized Design. Packed cell volume (PCV), red blood cell count (RBC), white blood cell count (WBC) and haemoglobin (HB) were determined using improved Neubaur Haemocytometer after dilution and Cyanomethamoglobin methods. Dressing percentage for the carcass yield were also obtained. Eosinophil counts and Uric acid were significantly ($p<0.05$) higher in birds administered 75 g of LBFE/ litre of water 5days/week. At the finisher phase, Packed cell volume was significantly ($P<0.05$) higher (32.25) in birds administered 75 g of LBFE/ litre of water 5days/week, while birds administered 125 g of LBFE/ litre of water as antibiotic replacement had significantly ($P<0.05$) higher (5.00) red blood cell count. The thigh and liver of birds administered LBFE 3 days/week had the highest values ($P<0.5$) of 10.79% and 2.84%, respectively. The study concluded that administration of *Lagenaria breviflora* fruit extract to broilers enhanced the haematological indices as well as carcass yield of the birds.

Keywords: Haematological response; *Lagenaria breviflora*; carcass yield; Broilers

Running title: Effect of *Lagenaria breviflora* fruit extract on Haematology and Carcass yield of Broilers

Réponse hématologique et rendement de carcasse des poulets de chair administrés avec des extraits de fruits de courge tachetée (*Lagenaria Breviflora* Robert)



Résumé

Au fil des années, l'application thérapeutique des plantes a été utilisée dans la médecine folklorique pour explorer des alternatives naturelles aux promoteurs de croissance antibiotiques, en raison du développement de souches bactériennes résistantes et de la présence de résidus médicamenteux dans la viande de volaille. Cette étude a examiné les effets des concentrations et des fréquences d'administration de l'extrait de fruit de *Lagenaria breviflora* (EFLB) sur les indices hématologiques et le rendement en carcasse des poulets de chair. 384 oiseaux ont été répartis aléatoirement en six traitements : T1 : contrôle, T2 : 75 g de LBFE/litre pendant 3 jours/semaine, T3 : 75 g de LBFE/litre pendant 5 jours/semaine, T4 : contrôle, T5 : 125 g de LBFE/litre pendant 3 jours/semaine et T6 : 125 g de LBFE/litre pendant 5 jours/semaine. L'expérience a été conçue selon un plan factoriel 2×3 et les données ont été soumises à un plan complètement randomisé. Le volume des cellules packées (VCP), le nombre de globules rouges (GR), le nombre de globules blancs (GB) et l'hémoglobine (HB) ont été déterminés à l'aide d'un hémocytomètre Neubauer amélioré après dilution et des méthodes Cyanométhémoglobine. Le pourcentage de panage pour le rendement en carcasse a également été obtenu. Les comptages des éosinophiles et de l'acide urique étaient significativement ($p<0.05$) plus

élevés chez les oiseaux administrés avec 75 g de EFLB/litre d'eau pendant 5 jours/semaine. À la phase de finition, le volume des cellules packées était significativement ($P < 0.05$) plus élevé (32.25) chez les oiseaux administrés avec 75 g de EFLB/litre d'eau pendant 5 jours/semaine, tandis que les oiseaux administrés avec 125 g de EFLB/litre d'eau en remplacement des antibiotiques avaient un nombre significativement ($P < 0.05$) plus élevé (5.00) de globules rouges. Les cuisses et le foie des oiseaux administrés avec EFLB 3 jours/semaine avaient les valeurs les plus élevées ($P < 0.5$) de 10,79% et 2,84%, respectivement. L'étude a conclu que l'administration d'extrait de fruit de *Lagenaria breviflora* aux poulets de chair améliorait les indices hématologiques ainsi que le rendement en carcasse des oiseaux.

Mots-clés : Réponse hématologique ; *Lagenaria breviflora* ; Rendement en carcasse ; Poulets de chair

Introduction

The poultry industry has become an important economic activity in many countries however, immunosuppression has posed a major challenge because affected flocks are susceptible to secondary infections, respond poorly to vaccines and do not perform as well as non-affected birds (Calcagni and Elenkov, 2006). The fruit of *Lagenaria breviflora* is widely used in folklore medicine in West Africa. It is used in animal production due to its antimicrobial property. (Ekunseitan *et al.*, 2023). Farmers employ the fruit extract of the plant for the treatment of Newcastle disease and coccidiosis in animals, especially poultry (Sonaiya, 2004). Researchers have validated the use of the plant as an anti-implantation agent (Elujoba *et al.*, 1985), abortifacient (Elujoba and Hymete, 1986), miracidicide and cercaricide (Ajayi *et al.*, 2002), as well as an antibacterial agent with broad spectrum activity (Tomori *et al.*, 2007). Adeyemi *et al.* (2017) observed that the presence of active ingredients and their biological activities in the fruit of *Lagenaria breviflora* makes it a prospective alternative to synthetic drugs in preventing and treating diseases in both man and animals. The active constituents in the leaves, stem, seed, root and fruit of these medicinal plants are highly effective in combatting different diseases and also aiding digestion which would ultimately improve the performance of recipients (Ashayerizadeh *et al.*, 2009). Compared to synthetic antibiotics or inorganic chemicals, products that are plant derivatives have been found to be less toxic, residue free, ideal feed additives for animal nutrition and most importantly they are natural (Wang and Bourne, 1998). Herbal plants are safe and economical. A properly prescribed amount of plant extract intake has numerous health benefits. This can be particularly seen in its application in animal feeding such as botanical additives in poultry feeds where results such as

digestive secretions, immune stimulation, antibacterial coccidiostatic, antiviral or anti-inflammatory activity in birds have been recorded (Cross *et al.*, 2007). However, Kalimuthu (2010) reported that though extract of plant which form the basis for all traditional system of medicine have been used for the treatment of various diseases, the lethal effects of these extracts maybe preceded by clinical signs of toxicity such as salivation, changes in animal eye colour, decreased respiratory rate and motor activity (Balogun *et al.*, 2014).

Haematological indices are useful parameters that can be employed to assess the toxic potentials of plant extracts in living systems (Sunmonu and Oloyede, 2010). Balogun *et al.*, (2014) opined that haematological parameters provide information regarding the status of bone marrow activity and hemolysis. Changes in haematological parameters can be used to determine the extent of deleterious effect of foreign compounds including plant extracts on the blood constituents of an animal (Ashafa *et al.*, 2009) because haematological indices are used as a base line in determination of damage to blood cells and in evaluation of responses of animal to therapy (Awotuyi, 1999).

Overall, it has been observed that serum biochemical and haematological parameters are improved by inclusion of phytobiotics in broiler diets (Al-Kassie, 2010). Serum biochemical and haematological parameters are reliable indicators of health status of animals, and may have important roles in diagnosis, prognosis, and treatments of poultry diseases. For example, alanine transaminase (ALT) and aspartate aminotransferase (AST) are considered diagnostic enzymes for liver diseases (Al-Jaff, 2011).

The study on administration of dosage gives an insight to safe use of *Lagenaria breviflora* fruit extract in poultry. Previous study had been carried out using laying birds focused on the effect on laying performance (Ekunseitan *et al.*,

2017), toxicology and biochemical (Balogun *et al.*, 2014) as well as phytochemical (Adeyemi *et al.*, 2017). Based on the findings above, this research was aimed at determining the effect of different concentration and frequency of administration, of *Lagenaria breviflora* fruit on the Haematological, serological and carcass yield of broiler chickens.

Materials and Methods

Experimental site

The field experiment was carried out at the Poultry Unit, Directorate of University Farms (DUFARMS), Federal University of Agriculture, Abeokuta, Ogun State, Nigeria. The area is situated in the South Western part of Nigeria. It is a region of 70m above the sea level of latitude 7° 5' to 7° 8'N and longitude 3°11'E. (Google Earth, 2021).

Sourcing and preparation of *Lagenaria breviflora* fruit extract

Lagenaria breviflora fruits were purchased from a local market in Abeokuta, Ogun State, Nigeria. The fruit sample was identified and authenticated as *Lagenaria breviflora* at the Department of Botany, Federal University of Agriculture Abeokuta. The whole fruit of *Lagenaria breviflora* was weighed, rinsed with distilled water and chopped into smaller sizes. The fruits were properly washed, weighed (75 g and 125 g) into two separate containers of

water at room temperature, respectively and soaked for 72 hours, while being stirred vigorously at 12-hour intervals. After soaking for 72 hours, it was sieved with muslin cloth to obtain a clear solution.

Experimental design

A total of 384 male 1-day- old Arbor acres birds were used for this experiment. Birds were fed with commercial broiler starter feed with crude protein of 23% and metabolizable energy of 2800Kcal/kg at the brooding phase while the broiler finisher feed had crude protein of 18% CP and metabolizable energy of 3000Kcal/kg, fed at the finishing stage. Feed and water containing the appropriate doses of *Lagenaria breviflora* fruit extracts were provided *ad libitum* throughout the experiment. All vaccinations and medications were administered and supplementation was carried out when and where necessary. After 2 weeks of brooding, the birds were balanced for weight and randomly distributed into experimental treatments. The birds were randomly distributed into 8 treatments; each treatment was assigned 48 birds. The treatments were further divided into 4 replicates with each replicate containing 12 birds. The experimental design is as shown in Table 1.

Table 1: Experimental design

Treatment	No of Birds/treatment	No of Replicates	No of Birds per replicate
T1 (control)	48	4	12
T2 (75 g/L/3days/week)	48	4	12
T3 (75 g/L/5days/ week)	48	4	12
T4 (control)	48	4	12
T5 (125 g/L/3days/week)	48	4	12
T6 (125 g/L/5days/week)	48	4	12

Where:

T1: Water and synthetic antibiotic as expected for routine medication (control for 75g/L), T2: 75 g of *Lagenaria breviflora* fruit extract in 1 litre of drinking water for three consecutive days/week, T3: 75 g of *Lagenaria breviflora* fruit extract in 1litre of drinking water for five consecutive days/week, T4: Water and synthetic antibiotic as expected for routine medication (control for 125g/L), T5: 125 g of *Lagenaria breviflora* fruit extracts in one litre of drinking water for three consecutive days/week, T6: 125 g of *Lagenaria breviflora* fruit extract in 1litre of drinking water for five consecutive days /week.

Phytochemical screening of the *Lagenaria breviflora* fruit extract

Phytochemical screening of the *Lagenaria breviflora* fruit extract in cold water was carried out using standardized procedures to determine the presence of tannin, reducing sugar, cardiac glycoside, alkaloid, saponin, flavonoid and phenol.

Haematological analysis of broilers

Blood samples were collected at the 2nd and 6th week of the experiment from two broiler chickens per replicate using syringe and needle through the wing vein. Samples were collected into a set of sterilized tubes containing Ethylene Diamine Tetra-acetic Acid (EDTA) labeled bottles, for the analysis of haematological

parameters. Packed cell volume (PCV), red blood cell count (RBC), white blood cell count (WBC) and haemoglobin (HB) were determined using improved Neubaur Haemocytometer after dilution and Cyanomethamoglobin methods respectively as described by Dacie and Lewis (1991). Mean corpuscular volume (MCV), Mean Corpuscular Haemoglobin (MCH) and Mean Corpuscular Haemoglobin Concentration (MCHC) were determined by the method of Hyduke (1995).

Serum biochemical analysis of broilers

Blood samples were collected at the 2nd and 6th week of the experiment from two broiler chickens per replicate using syringe and needle through the wing vein and kept inside plain sample bottles. Serum urea and creatinine level were determined using spectrophotometric methods described by Coles (1986). The activities of hepatic marker enzymes were determined in the serum. Alkaline phosphatase. Alanine transaminase (ALT) and aspartate transaminase (AST) activities were determined using the method described by Reitman and Frankel, (1957).

Antibody Titre

Haemagglutination inhibition tests for Newcastle disease (ND) antibodies: Blood samples were collected at the 1st, 3rd, 5th, 6th and 8 week of age for determination of antibody titre against Newcastle Disease and Blood samples were left without anticoagulant to clot. The serum was separated by centrifugation at 3000 rpm for 10 min. Microtechnique of haemagglutination inhibition test was done according to Takatasy (1955). Geometric mean titer (GMT) was calculated according to Brugh (1978).

Carcass yield

At the 42nd day of the experiment, two birds of average weight per replicate were selected. The birds were fasted for 12 hours to empty their stomach to prevent carcass contamination by gut content but they had access to water to avoid dehydration. Birds were slaughtered and dressed following conventional procedure by severing the carotid artery and jugular vein. The birds were bled completely followed by scalding in water at 60 °C. The live weight, plucked weight and dressed weight were recorded.

$$\text{Dressing percentage (\%)} = \frac{\text{Dressing Weight (g)} \times 100}{\text{Live Weight (g)}}$$

Organs which include gizzard, liver, heart and the intestines were weighed. Cut parts of the birds such as the breast, back, thigh, drum stick, head, wings and shanks were weighed and expressed as percentages of the live weight.

$$\text{Cut part percentage (\%)} = \frac{\text{Cut part weight (g)} \times 100}{\text{Live Weight (g)}}$$

Statistical analysis

Data obtained were subjected to 2 x 3 factorial Analysis of Variance in a Completely Randomized Design. Significant differences among treatment means were separated using Duncan's Multiple Range Test with the aid of SPSS (2021) package at 5% probability Frequency of administration.

Results

Phytochemical Screening of the *Lagenaria breviflora* fruit extract

The results of the phytochemical screening show the presence of metabolites both at the 75g/litre and 125g/litre preparation as shown in Table 2.

Table 2: Phytochemical screening of *Lagenaria breviflora* fruit extract

Parameters (mg/100g)	75 g of LBFE/L of water
Tannins	20.95
Reducing sugars	30.26
Cardiac glycoside	36.73
Alkaloids	44.12
Saponins	40.48
Flavonoids	33.94
Phenol	28.81

Main effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at starter phase

The main effects of concentration and frequency of administration of LBFE on haematological parameters of broiler chickens at starter phase are showed in Table 3. Lymphocyte and eosinophil counts differed significantly ($p < 0.05$) as influenced by concentration of LBFE. Lymphocyte count was higher ($p < 0.05$) in birds administered 125 g/L of the extract. However, eosinophil count was significantly ($p < 0.05$) higher (2.81%) in birds administered 75 g of LBFE /L of water. The effect of frequency of administration of LBFE influenced only white blood cell count with

significantly ($p < 0.05$) highest count (109125 g/L) observed in birds in the control and lowest (36375 g/L) in birds given antibiotic replacement.

Table 3: Main Effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at starter phase

Parameters	Concentration				Frequency of administration				
	75 g/L	125 g/L	SEM	P value	Control	3days/wk	5days/wk	SEM	P value
Packed Cell Volume (%)	35.03	34.25	1.23	0.655	34.38	35.88	34.56	1.74	0.851
White Blood Cell count (g/L)	73125	66125	10194	0.629	109125 ^a	75250 ^{ab}	57750 ^{ab}	14416	0.006
Red Blood Cell Count ($10^{12}/L$)	4.47	4.91	0.29	0.290	4.50	4.88	4.94	0.41	0.762
Heterophil (%)	58.91	56.44	2.46	0.482	58.25	57.00	56.19	3.48	0.929
Lymphocyte (%)	30.09 ^b	36.97 ^a	2.11	0.025	35.63	30.69	36.06	2.98	0.484
Monocyte (%)	5.03	5.44	0.59	0.628	4.75	4.44	4.56	0.83	0.072
Eosinophil (%)	2.81 ^a	1.50 ^b	0.37	0.014	1.63	1.25	2.94	0.52	0.057
Basophil (%)	0.31	0.28	0.15	0.887	0.00	0.38	0.25	0.22	0.329
Heterophil : Lymphocyte	1.96	1.53			1.63	1.86	1.56		

^{ab} Means on the same row having different superscripts are significantly ($p < 0.05$) different wk week

Effects interaction between of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at starter phase

Table 4 shows the interaction between concentration and frequency of administration of LBFE on haematological parameters of

broiler chickens at starter phase. All parameters measured except eosinophil count were not significantly ($p > 0.05$) different across treatments. Eosinophil counts were highest (4.50%) in birds administered 75 g of LBFE / litre of water 5days/week and lowest (1.00%) in birds administered 125 g of LBFE / litre of water 3days/week.

Table 4: Effects of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at starter phase

Parameters	T1	T2	T3	T4	T5	T6	SEM	P value
Packed Cell Vol. (%)	34.38	36.38	34.13	34.38	35.38	35.00	2.46	0.875
WB C (g/L)	109125	73625	79125	109125	76875	36375	20387	0.549
R B C $\times 10^{12}/L$	4.50	4.50	4.75	4.50	5.25	5.13	0.58	0.921
Heterophil (%)	58.25	55.88	59.63	58.25	58.13	52.75	4.93	0.767
Lymphocytes (%)	35.63	24.88	31.25	35.63	36.50	40.88	4.22	0.539
Monocytes (%)	4.75	4.63	4.62	4.75	4.25	4.50	1.18	0.697
Eosinophils (%)	1.63 ^{ab}	1.50 ^{ab}	4.50 ^a	1.63 ^{ab}	1.00 ^b	1.38 ^{ab}	0.73	0.161
Basophil (%)	0.00	0.63	0.00	0.00	0.13	0.50	0.31	0.449

Heterophil:lymph
ocyte

1.63 2.25 1.91 1.63 1.59 1.29

^{ab} Means on the same row having different superscripts are significantly ($p < 0.05$) different
WBC - White Blood Cell, RBC - Red Blood Cell

Main effect of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at starter phase

The main effects of concentration and frequency of administration of LBFE on serum biochemical parameters of broiler chickens at starter phase are revealed in Table 5. All

parameters measured were not significantly ($p > 0.05$) affected by concentration of LBFE. However, the effect of frequency of administration of LBFE significantly ($p < 0.05$) affected uric acid. Uric acid was significantly ($p < 0.05$) highest (35.19mg/dL) in birds fed LBFE 5days/week and lowest (14.38mg/dL) in birds in the control.

Table 5: Main Effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at starter phase

HN	Concentration				Frequency of administration				
Parameters	75 g/L	125 g/L	SEM	P value	Control	3 days/week	5 days/week	SEM	P value
Uric acid (mg/dL)	22.06	23.67	3.46	0.744	14.38 ^b	20.78 ^{ab}	35.19 ^a	4.90	0.030
Creatinine (mg/dL)	1.919	1.413	0.502	0.479	0.863	1.388	2.025	0.710	0.444
ALT (U/L)	16.06	18.97	4.49	0.649	10.13	21.94	18.87	6.34	0.586
AST (U/L)	21.62	20.06	6.08	0.856	8.75	34.00	22.19	8.59	0.231

^{ab} Means on the same row having different superscripts by factor are significantly ($p < 0.05$) different

Effect of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at starter phase

The effect of interaction between concentration and frequency of administration of LBFE on serum biochemical parameters of broiler chickens at starter phase is presented in Table

6. All parameters measured except Uric acid were not significantly ($p > 0.05$) different across treatments. Uric acid was significantly ($p < 0.05$) highest (50.75mg/dL) in birds administered 75 g of LBFE /L of water 5 days/week while lowest values (11.50mg/dL) was recorded in birds administered 75 g of LBFE /L of water as antibiotic replacement which was similar to the value obtained from the other groups.

Table 6: Effects of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at starter phase

Parameters	T1	T2	T3	T4	T5	T6	SEM	P value
Uric acid (mg/dL)	14.38 ^b	11.63 ^b	50.75 ^a	14.38 ^b	29.92 ^{ab}	19.62 ^b	6.93	0.002
Creatinine (mg/dL)	0.86	0.63	2.87	0.86	2.15	1.17	1.00	0.298
ALT (U/L)	10.13	8.63	30.12	10.13	35.25	7.62	8.97	0.064
AST (U/L)	8.8	26.5	38.6	8.8	41.5	5.8	12.2	0.196

^{ab} Means on the same row having different superscripts by factor are significantly ($p < 0.05$) different

Main effect of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at finisher phase

The main effects of concentration and frequency of administration of LBFE on

haematological parameters of broiler chickens at finisher phase is shown in Table 7. Effect of concentration of the extract had no significant ($p > 0.05$) effect on all the measured parameters. However, effect of frequency of administration of LBFE influenced significantly ($p < 0.05$)

Packed cell volume and Eosinophil count. Packed cell volume was significantly ($p<0.05$) increased (29.87%) with the frequency of

administration of the extract while eosinophil count was reversed other.

Table 7: Main Effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at finisher phase

Parameters	Concentration				Frequency of administration				
	75 g/L	125 g/L	SEM	P value	Control	3days/ week	5days/ week	SEM	P value
Packed Cell Volume (%)	26.56	26.84	1.11	0.858	22.13 ^b	27.62 ^{ab}	29.87 ^a	1.57	0.008
White Blood Cell (g/L)	18000	21937	4311	0.521	16875	27500	20562	6097	0.486
Red Blood Cell ($\times 10^{12}/L$)	3.688	3.875	0.147	0.370	3.375	3.500	4.125	0.207	0.016
Heterophil (%)	62.16	63.81	2.18	0.593	63.88	58.25	63.31	3.08	0.300
Lymphocytes (%)	30.44	28.06	2.08	0.423	26.13	35.06	30.06	2.95	0.102
Monocytes (%)	3.531	4.656	0.404	0.054	5.000	3.563	3.938	0.571	0.318
Eosinophils (%)	3.531	3.281	0.424	0.678	4.875 ^a	2.813 ^{ab}	2.313 ^b	0.599	0.022
Basophil (%)	0.187	0.188	0.116	1.000	0.125	0.125	0.250	0.164	0.900
Heterophil :Lymphocyte	2.04	2.27			2.44	1.66	2.11		

^{ab} Means on the same row having different superscripts by factor are significantly ($p<0.05$) different

Effects of interaction between concentration and frequency of administration of Lagenaria breviflora fruit extract on haematological parameters of broiler chickens at finisher phase

Interaction between concentration and frequency of administration of LBFE on haematological parameters of broiler chickens at finisher phase is presented in Table 8. Packed cell volume and Red blood cell count were significantly ($p<0.05$) different across

treatments. Packed cell volume was significantly ($P<0.05$) highest (32.25%) in birds administered 75 g of LBFE / litre of water 5days/week, while lowest value (22.13%) was obtained in birds in the control group administered 75 g of LBFE / litre of water, and birds in the control group. Birds administered 125 g of LBFE / litre of water as antibiotic replacement had significantly ($P<0.05$) highest (5.00L) red blood cell count.

Table 8: Effects of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on haematological parameters of broiler chickens at finisher phase

Parameters	T1	T2	T3	T4	T5	T6	SEM	P value
Packed Cell Volume (%)	22.13 ^b	29.25 ^{ab}	32.25 ^a	22.13 ^b	26.00 ^{ab}	27.50 ^{ab}	2.21	0.013
White Blood Cell (g/L)	16875	28250	7750	16875	26750	33375	8622	0.226
Red Blood Cell ($\times 10^{12}/L$)	3.375 ^b	3.875 ^{ab}	4.250 ^{ab}	3.375 ^b	3.125 ^b	4.000 ^{ab}	0.293	0.001
Heterophil (%)	63.88	54.88	65.25	63.88	61.63	61.38	4.36	0.645
Lymphocytes (%)	26.13	38.25	29.37	26.13	31.88	30.75	4.17	0.763
Monocytes (%)	5.000	3.000	2.375	5.000	4.125	5.500	0.808	0.214
Eosinophils (%)	4.875	3.500	2.250	4.875	2.125	2.375	0.847	0.754
Basophil (%)	0.125	0.000	0.500	0.125	0.250	0.000	0.231	0.331

Heterophil :

Lymphocyte 2.44 1.43 2.22 2.44 1.93 2.00

^{ab} Means on the same row having different superscripts are significantly ($p < 0.05$) different**Main effect of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at finisher phase**

The main effects of concentration and frequency of administration of LBFE on serum biochemical parameters of broiler chickens at finisher phase are shown in Table 9. All parameters measured were not significantly ($p > 0.05$) affected by concentration of LBFE. However, the effect of Frequency of

administration of LBFE significantly ($p < 0.05$) affected uric acid and ALT. Uric acid was significantly ($p < 0.05$) highest (32.81mg/dL) in birds fed antibiotic replacement and lowest values (17.88 and 20.63mg/dL) were recorded in birds in the control and birds administered LBFE 3days/week, respectively. ALT was significantly ($p < 0.05$) highest (23.25U/L) in birds in the control and lowest (10.56U/L) in birds administered LBFE 5days/week.

Table 9: Main Effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at finisher phase

Parameters	Concentration				Frequency of administration				SEM	P value
	75 g/l	125 g/l	SEM	P value	Control	3days/wk	5days/wk	SEM		
Uric acid (mg/dL)	22.59	25.56	2.24	0.353	17.88 ^b	20.63 ^b	25.00 ^{ab}	3.17		0.009
Creatinine (mg/dL)	0.988	1.337	0.160	0.128	0.850	1.150	1.006	0.226		0.087
ALT (U/L)	15.47	12.41	2.38	0.368	23.25 ^a	10.88 ^{ab}	10.56 ^b	3.37		0.024
AST (U/L)	11.28	9.25	1.76	0.417	10.25	8.13	13.38	2.49		0.489

^{ab} Means on the same row having different superscripts by factor are significantly ($p < 0.05$) different**Effect of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at finisher phase**

The interaction between concentration and frequency of administration of LBFE on serum biochemical parameters of broiler chickens at finisher phase shown on Table 10 revealed all the measured parameters were not significantly ($p > 0.05$) different across treatments.

Table 10: Effects of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on serum biochemical parameters of broiler chickens at finisher phase

Parameters	T1	T2	T3	T4	T5	T6	SEM	P value
Uric acid (mg/dL)	17.88	15.00	24.87	17.88	26.25	25.12	4.49	0.523
Creatinine (mg/DL)	0.850	1.013	0.925	0.850	1.287	1.087	0.320	0.460
ALT (U/L)	23.25	12.63	14.50	23.25	9.13	6.63	4.77	0.843
AST (U/L)	10.25	8.63	13.25	10.25	7.63	13.50	3.51	0.667

Main effect of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on Antibody titre to Newcastle Disease vaccination of broiler chickens

The main effects of concentration and frequency of administration of LBFE on immuno-modulatory of broiler chickens are presented in Table 11 All measured parameters

were not significantly ($p > 0.05$) affected both concentration and frequency of administration of LBFE. Although, there was no significant difference however the mean titer of serum antibodies of Newcastle disease was higher in the birds that received 75 g of LBFE / litre of water at the starter phase compared to the control ($p > 0.05$).

Table 11: Main effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on Antibody titre to Newcastle Disease vaccination of broiler chickens

Parameters	75 g/L	Concentration			Control	Frequency of administration			
		125 g/L	SEM	P value		3days/w eek	5days/w eek	SEM	P value
Antibody titre at starter phase (log ₂ GMt)	37.9	18.2	12.4	0.265	13.8	17.0	12.6	17.5	0.075
Antibody titre at finisher phase (log ₂ GMt)	12.50	14.00	3.70	0.775	12.00	18.88	7.50	5.23	0.483

Effect of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on Antibody titre to Newcastle Disease vaccination of broiler chickens

The interaction between concentration and frequency of administration of LBFE on immuno-modulatory of broiler chickens is shown in Table 12. No significant ($p>0.05$) differences were observed across treatment means

Table 12: Effects of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on Antibody titre to Newcastle Disease vaccination of broiler chickens

Parameters	T1	T2	T3	T4	T5	T6	SEM	P value
Antibody titre at starter phase (log ₂ gmt)	13.8	13.8	20.7	13.8	20.3	4.5	24.7	0.418
Antibody titre at finisher phase (log ₂ gmt)	12.00	14.75	11.50	12.00	23.00	3.50	7.40	0.701

Main effect of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on Carcass characteristics of broiler chickens

The main effects of concentration and frequency of administration of LBFE on carcass characteristics of broiler chickens were presented in Table 13. All parameters except head were not significantly ($p>0.05$) affected by concentration of *Lagenaria breviflora* fruit

extract with significantly ($p<0.05$) higher (2.55%) head measured in birds administered 75 g of LBFE / litre of water. The effect of frequency of administration of LBFE influenced significantly ($p<0.05$) the liver portion with highest value (2.84%) obtained in birds administered LBFE 3days/week and lowest (2.35%) in birds administered antibiotic replacement.

Table 13: Main Effects of concentration and frequency of administration of *Lagenaria breviflora* fruit extract on carcass characteristics of broiler chickens

Parameters	Concentration				Frequency of administration				
	75 g/L	125 g/L	SEM	P value	Control	3days/ wk	5days/w k	SEM	P value
Live weight (kg)	1.75	1.70	0.04	0.377	1.76	1.71	1.67	0.05	0.629
Defeathered weight(kg)	1.60	1.56	0.03	0.416	1.63	1.58	1.51	0.05	0.391
Dressed weight (kg)	1.23	1.19	0.03	0.474	1.27	1.18	1.17	0.04	0.269
Dressing %	70.17	70.46	0.55	0.711	71.79	69.09	69.62	0.77	0.075
Head (%)	2.55 ^a	2.37 ^b	0.06	0.026	2.45	2.39	2.49	0.08	0.803
Shank (%)	4.12	3.91	0.10	0.137	4.08	4.10	3.86	0.14	0.632
Neck (%)	2.69	2.91	0.08	0.056	2.88	2.61	2.83	0.11	0.319
Breast (%)	19.74	19.74	0.34	0.992	20.17	20.14	18.67	0.48	0.094
Back (%)	14.95	14.78	0.24	0.613	15.41	14.16	15.17	0.34	0.057
Thigh (%)	9.89	9.86	0.17	0.902	9.52	9.51	10.24	0.24	0.041
Drumsticks (%)	9.49	9.58	0.15	0.682	10.03	9.25	9.30	0.21	0.045
Wings (%)	7.95	8.11	0.16	0.482	8.29	7.66	8.08	0.22	0.241
Liver (%)	2.60	2.59	0.07	0.980	2.64 ^{ab}	2.84 ^a	2.58 ^{ab}	0.10	0.013
Heart (%)	0.54	0.56	0.03	0.548	0.54	0.59	0.53	0.04	0.625
Gizzard (%)	1.88	1.85	0.05	0.749	1.85	1.84	1.79	0.07	0.356
Proventriculus (%)	0.45	0.47	0.02	0.421	0.46	0.50	0.44	0.03	0.330
GIT (%)	9.31	8.79	0.25	0.148	8.88	9.85	8.59	0.35	0.072

^{ab} Means on the same row having different superscripts are significantly (p<0.05) different

Effect of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on Carcass characteristics of broiler chickens

Interaction between concentration and frequency of administration of LBFE on carcass characteristics of broiler chickens is depicted in Table 14. Neck, thigh, liver and GIT were significantly (p<0.05) different across treatments. Neck was significantly (P<0.05) highest (3.40%) in birds administered 125 g of LBFE / litre of water 5days/week, while lowest values (2.26 and 2.22%) were obtained in birds administered 75 g of LBFE / litre of water 5days/week and birds administered 125 g of LBFE / litre of water 3days/week, respectively. Thigh was

significantly (P<0.05) highest (10.79%) in birds administered 125 g of LBFE / litre of water 5days/week and lowest (9.22%) in birds administered 125 g of LBFE / litre of water 3days/week. Liver was significantly (P<0.05) highest (2.95%) in birds administered 125 g of LBFE / litre of water 3days/week and lowest (2.23%) in birds administered 75 g of LBFE / litre of water as antibiotic replacement. The GIT was significantly (P<0.05) highest (10.68%) in birds administered 75 g of LBFE / litre of water 3days/week and lowest (8.35%) in birds administered 125 g of LBFE / litre of water 5days/week.

Table 14: Effects of interaction between concentration and frequency of administration of *Lagenaria breviflora* fruit extract on carcass characteristics of broiler chickens

Haematological Response and Carcass Yield Of Broiler Chickens Administered Spotted Pumpkin (*Lagenaria Breviflora* Robert) Fruit Extracts

Parameters	T1	T2	T3	T4	T5	T6	SEM	P value
Live weight (kg)	1.76	1.76	1.66	1.76	1.66	1.68	0.075	0.763
Defeathered weight (kg)	1.63	1.64	1.49	1.63	1.51	1.53	0.073	0.638
Dressed weight (kg)	1.27	1.22	1.13	1.27	1.14	1.20	0.057	0.401
Dressing percent (%)	71.80	69.35	67.83	71.80	68.84	71.41	1.09	0.088
Head (%)	2.45	2.58	2.66	2.45	2.23	2.33	0.12	0.314
Shank (%)	4.08	4.52	3.88	4.08	3.69	3.85	0.20	0.116
Neck (%)	2.88 ^{ab}	2.79 ^{ab}	2.26 ^b	2.88 ^{ab}	2.44 ^b	3.40 ^a	0.16	<0.001
Breast (%)	20.17	19.66	18.62	20.17	20.61	18.72	0.68	0.544
Back (%)	15.41	14.37	14.77	15.41	13.94	15.56	0.48	0.287
Thigh (%)	9.52 ^{ab}	9.81 ^{ab}	9.69 ^{ab}	9.52 ^{ab}	9.22 ^b	10.79 ^a	0.34	0.046
Drumsticks (%)	10.03	9.43	9.15	10.03	9.07	9.46	0.29	0.575
Wings (%)	8.29	7.64	7.89	8.29	7.67	8.27	0.31	0.922
Liver (%)	2.64 ^{ab}	2.72 ^{ab}	2.82 ^{ab}	2.64 ^{ab}	2.96 ^a	2.34 ^{ab}	0.15	0.054
Heart (%)	0.55	0.54	0.51	0.55	0.64	0.54	0.05	0.520
Gizzard (%)	1.85	1.91	1.72	1.85	1.77	1.88	0.09	0.443
Proventriculus (%)	0.46	0.47	0.41	0.46	0.53	0.47	0.04	0.510
GIT (%)	8.88 ^{ab}	10.68 ^a	8.84 ^{ab}	8.88 ^{ab}	9.03 ^{ab}	8.35 ^b	0.49	0.287

^{ab} Means on the same row having different superscript are significantly (p<0.05) different

Discussion

A reliable and fast means of assessing clinical and nutritional health status of animals include the use of blood analysis, because ingestion of

nutritional status (Togun and Oseni, 2005, Olabanji *et al.*, 2007). Haematological results in the starter phase of this study revealed that lymphocyte and eosinophil counts differed significantly as influenced by concentration of *Lagenaria breviflora* fruit extract. This is further confirming report of Ekunseitan *et al.* (2016) that *Lagenaria breviflora* Robert (Spotted Pumpkin) possesses relevant ethno-veterinary properties in the prevention and treatment of rural poultry health management. The same trend was observed in the finisher phase where birds in the control had higher eosinophil count than birds administered *Lagenaria breviflora* fruit extract for 5days/week. Although the birds were in a good state of health during this study, however, elevated WBC and eosinophil counts can be an indicator of disease or infection. It shows that the birds are producing more antibodies to fight off infection. A number of in vitro and animal studies have shown that bioactive compounds from plants increase immunologic activity by increasing phagocytosis (Geetha *et al.*, 2012; Lillehoj *et al.*, 2018). Phagocytosis by granulocytes is the first and major defense mechanism against invasion of bacteria, fungi,

bioactive components have measurable effects on blood composition (Church *et al.*, 1984; Maxwell *et al.*, 1990) and may be considered as appropriate measure of long term

and parasites. Nonetheless, the values obtained are within the normal reference range for domestic chickens (Jain, 1986). The higher Packed cell volume in treatments administered *Lagenaria breviflora* fruit extract observed in finisher broilers in this study can be due to their inability to cause haemolysis inherent from the anti-inflammatory potential inherent in *Lagenaria breviflora*. Similar results were reported by Ekunseitan *et al.* (2017) where packed cell volume and red blood cell counts were highest in laying birds administered *Lagenaria breviflora* fruit extract.

Biochemical components are sensitive to elements of toxicity in feeds and plant extract and are particularly valuable in monitoring toxicity especially with feed constituents that affect the formation of blood (Oyawoye and Ogunkunle, 1998). The results of serum biochemical indices measured in this study revealed higher uric acid in treatments at both starter and finisher phases and it agrees with the findings of Ekunseitan *et al.* (2017) on significantly higher serum uric acid in laying birds administered *Lagenaria breviflora* extract than birds in the control. The kidney has been recognized as the main regulator of serum

uric acid and the excretion of renal uric acid is determined by the balance of urate re-absorption and re-secretion (Bobulescu and Moe, 2012). Therefore, serum uric acid frequency of administration could be regarded as a therapeutic target for the prevention of thromboembolic events and may be an indicator of antithrombotic therapy decision (Dogan *et al.*, 2016). In addition, ALT was higher in birds in the control than in birds administered *Lagenaria breviflora* fruit extract thus, indicating the liver health was in the best condition in birds in this treatment. Plant extracts have been reported by previous reports by Muhammad *et al.* (2020) to contain certain compounds that reduce hepatic damage. These plants are rich source of alkaloids, flavonoids, diterpenoid, tannins, lipids, sterols etc. with anti-microbial and hepatic protective effects (Mangwani *et al.*, 2019).

Furthermore, the antibody titre against Newcastle disease effects of *Lagenaria breviflora* fruit extract in broiler chickens at starter and finisher phases as investigated in this present study is negligible. Similar reports were presented by Faluyi and Agbede (2018) who observed no significant differences when investigating the immuno-modulatory effects of aqueous leaf extracts of *Moringa oleifera* in broiler chickens in response to Newcastle disease vaccinations.

The antibody titre values reduced as the WBC reduced in both concentrations 75 g and 125 g of *Lagenaria breviflora* fruit extract per litre of water at the starter and finisher phases. Also the titre values reduced as the concentration increased. There is consistency in the trend of immune response as evident in lymphocyte observed at both 75 g and 125 g of the extract per litre of water in the starter phase. The increase in B-lymphocyte could be attributed to the improved immune response of the birds receiving *Lagenaria breviflora* extract. This study agrees with Ali *et al.* (2018) who observed Titer of serum antibodies of Newcastle disease was maximum in treatment supplemented with CLA+HMB compared to the control. The effect of *Lagenaria breviflora* extract observed in the current study was a significant increase in WBC and lymphocytes respectively which showed an improvement in immune system. The present study is in agreement with those obtained by Soltan (2009) that enzyme had no effect of HI

titer on broiler chicks when compared with the chicks fed on the same diet containing different levels of palm kernel cakes without enzyme supplementation.

The response of the birds to *Lagenaria breviflora* may pose some stress in the body system of the birds thereby increasing the production of white blood cell count and lymphocyte. Moreover, the effect of the extract may have modulated the immune response of the birds. The increased lymphocyte count may be reflection of higher activity of humeral immune responses in chicks. Ali *et al.* (2018) reported that increasing in lymphocyte populations may be indicative of higher activity of humeral immune responses in chicks fed HMB and CLA supplemented diets.

The significant differences across treatments observed in carcass traits such as neck, thigh, liver and GIT as influenced by the interaction between concentration and frequency of administration of LBFE agrees with previous studies by Jamroz and Kamel (2002) and Alçiçek *et al.* (2003) that significant effect of herbal extract was recorded on carcass characteristics of broiler chickens. Al Kassie (2010) also recorded positive effect in liver yield of broiler administered different frequency of administration of peppermint. In contrast, Hernandez *et al.* (2004) and Amouzmehr *et al.* (2012) reported that use of herbal extracts had no significant effect on carcass characteristics. These disparities could result from differences in plant extract used and frequency of administration.

Conclusion

This study has been able to show that the administration of *Lagenaria breviflora* fruit extract to broiler birds can potentiate their haematological indices as well as improve the carcass yield. This invariable implies higher yield and more availability of meat to consumers.

References

- Adeyemi, M. A., Ekunseitan, D. A., Abiola, S. S., Dipeolu, M. A., Egbeyale, L. T. and Sogunle, O. M. 2017. Phytochemical Analysis and GC-MS Determination of *Lagenaria breviflora* R. Fruit. *International Journal of Pharmacognosy and Phytochemical Research* 9 (7): 1051-1056

- Ajayi, G.O., Awajo, N.C. and Abulu, L.E. 2002.** Themiracidial and cercaricidal activity of the methanolic extract of *Lagenaria breviflora* Robert family Cucurbitaceae fruit in *Schistosoma mansoni*. *Nigerian Quarterly Journal of Hospital Medicine*, 12: 57-59.
- Alcicek, A., Bozkurt, M. and Abuk, M. C. 2004.** The effect of a mixture of herbal essential oils, an organic acid or a probiotic on broiler performance. *South African Journal of Animal Sciences* 34(4): 217-222.
- Alçiçek, A., Bozkurt, M. and Çabuk, M. 2003.** The effects of an essential oil combination derived from selected herbs growing wild in turkey on broiler performance. *South African Journal of Animal Sciences* 33:89-94.
- Ali, A.S., Vahid, K., Ali, R. N. and Pouya, Z. 2018.** Immune response, blood parameters and growth performance in broiler fed reduced protein diet supplemented with β -hydroxy- β -methyl butyrate and conjugated linoleic acid. *Acta Scientiarum Animal Sciences* 40(1) e38000.
- Al-Jaff F.K. 2011** Effect of coriander seeds as diet ingredient on blood parameters of broiler chicks raised under high ambient temperature. *International Journal Poultry Science* 10: 82-86.
- Al-Kassie G.A.M. 2010** The effect of thyme and cinnamon on the microbial balance in gastro intestinal tract on broiler chicks. *International Journal Poultry Science* 9: 495-498.
- Amouzmehr, A., Dastar, B., Nejad, J. G., Sung, K. I., Lohakare, J. and Forghani, F. 2012.** Effects of Garlic and Thyme Extracts on Growth Performance and Carcass Characteristics of Broiler Chicks. *Journal of Animal Science and Technology* 54(3):185-190
- Ashafa, A.O.T., Yakubu, M.T., Grierson, D.S. and Afolayan, A.J. 2009** Effects of aqueous leaf extract from the leaves of *Chrysocoma ciliate* L. on some biochemical parameters of Wistar rats. *African Journal of Biotechnology* 8: 1425-1430.
- Ashayerizadeh, A., Dastar, B., Shams Shargh M., Ashayerizadeh, A., Rahmatnejad, E., and Hessaini, S.M.R. 2009** Use of Garlic (*Allium sativum*), Black Cumin seeds (*Nigella sativa* L.) and Wild Mint (*Mentha longifolia*) in broiler chicken diets. *Journal of Animal Veterinary Advances* 9:1860-1863.
- Awotuyi, E. K. 1999.** A study of baseline haematological value of domestic and commercial chicken in Ghana, *Bullettin Animal production Africa* 38:453-458.
- Balogun, M.E., Ajayi, A.F., Oji, J.O., Besong, E.E., Finbarrs-Bello, E. and Folawiyo, M.A. 2014.** Toxicological and biochemical studies of ethanolic fruit extract of *Adenopus breviflorus* (*Lagenaria breviflora* Robert) in male albino wistar rats. *American Journal of Phytomedicine and Clinical therapeutics* 2(9): 1112-1123.
- Bobulescu, I.A. and Moe, O.W. 2012.** Renal transport of uric acid: evolving concepts and uncertainties. *Advanced Chronic Kidney Disease* 19: 358-371.
- Calcagni, E. and Elenkov, I. 2006** Stress system activity, innate and T helper cytokines, and susceptibility to immune-related diseases. *Annals of the New York Academy of Science* 1069: 62-76.
- Church, J. P., Judd, J. T., Young, C. W., Kebay, J. L. and Kim, W. W. 1984.** Relationships among diet constituents and specific serum clinical components of subjects eating self selected diets. *South African Journal of Clinical Nutrition* 40:1338-1344.
- Coles, E.H. 1986.** Veterinary clinical pathology 4th ed E.B. Saunder Company, Philadelphia and London. 220 - 239
- Cross, D.E., McDevitt, R.M., Hillman, K., and Acamovic, T., 2007.** The effect of herbs and their associated essential oils on performance, dietary digestibility and gut microflora in chickens from 7 to 28 days of age. *British Poultry Science* 4:496-506.
- Dogan, M., Uz, O. and Bicakci, B. 2016.** Elevated uric acid Frequency of administration: the chicken or the egg? *Journal of Geriatric Cardiology*. 13(4): 367 – 368.

- Ekunseitan, D. A., M. O. Akinola, W. O. Adegun, E. O. Akinwunmi and T. M. Omoseyin 2023.** Quality attributes and antioxidant activities of meat of broiler chickens administered aqueous *Lagenaria breviflora*. *Agricultura tropica et subtropica*, 56: 74–80
- Ekunseitan, D.A., Yusuf, A. O., Olayinka, O.A., Ayoola, A.A. and Adegbenjo, A.A. 2016.** Comparative study of two plants (*Lagenaria breviflora* and *Petiveria alliacea*) and their phytobiotic potential in poultry health. *Nigerian Journal of Animal Production* 43(1): 289-298.
- Ekunseitan, D.A., Abiola, S.S., Oluwatosin, O.O., Sogunle, O.M., Adeleye, O.O., Egbeyale, L.T. and Iyasere, O.S. 2017.** Health status of laying birds administered extracts of *Lagenaria breviflora* managed under two housing systems. *Journal of Animal Science* 97: 154–164
- Elujoba, A.A., Olagbende. S.O. and Adesina, S. K. 1985.** Anti-implantation activity of the fruit of *Lagenaria breviflora* Robert. *Journal of Ethnopharmacology* 13: 281-288.
- Elujoba A. A. and Hymete A. 1986.** Abortifacient activity of the fruit pulp of *Lagenaria breviflora*. *Fitotera*. 57: 97-101.
- Faluyi, O.B. and Agbede, J.O. 2018.** Immuno-Modulatory Activity of Aqueous Leaf Extract of *Moringa Oleifera* in Broiler Chickens. *International Journal of Environment, Agriculture and Biotechnology (IJEAB)* 3(1): 49 – 54
- Geetha, R.V., Lakshmi, T. and Roy, A. 2012.** A review on nature's immune boosters. *International Journal of Pharmaceutical Sciences Review and Research* 13:43–52
- Google Earth, (2021).** <https://earth.google.com/web> Retrieved 21st March, 2021.
- Hernandez, F., Madrid, J., Garcia, V., Orengo, J. and Megias, M. D. 2004.** Influence of two plants extracts on broiler performance, digestibility and digestive organ size. *Poultry Science* 83:169-174.
- Hyduke, R. R. 1995.** Chemical Biochemistry Laboratory Manual. The University of Iowa Medical Technology Programme Iowa City, U.S.A.
- Jain, NC 1986,** *Schalm Veterinary Haematology* (4th edition). Lea and Febiger, Philadelphia 285.
- Jamroz, D. and Kamel, C. 2002.** Plant extracts on immunity, health and performance. *Journal of Animal Science* 80:41.
- Kalimuthu K., S. Vijayakumar and R. Senthilkumar. 2010.** Antimicrobial Activity of the Biodiesel plant, *Jatropha curcas* L. *International Journal of Pharmacology and BioSciences* 1(3):1-5
- Lillehoj, H., Liu, Y., Calsamiglia, S. 2018** Phytochemicals as antibiotic alternatives to promote growth and enhance host health. *Veterinary Research* 49: 76.
- Mangwani, N., Singh, P.K. and Kumar, V. 2019.** Medicinal plants: Adjunct treatment to tuberculosis chemotherapy to prevent hepatic damage, *Journal of Ayurveda and Integrative Medicine* 11(4): 522-528
- Maxwell, M. H., Robertson, G. W., Spences and McCongrouodala, C. C. 1990** Composition of haematological values in restricted and ad-libitum feed domesticated fowls:- RBC Characteristics. *British Poultry* 60:1474-1484.
- Muhammad, A. B., Arifah, A. K., Teck, C. L., Saleha, A. A., Annas, S., Ubodullah K. and Sherifat, B. I. 2020** Effects of Inclusion of Different Doses of *Persicaria odorata* Leaf Meal (POLM) in Broiler Chicken Feed on Biochemical and Haematological Blood Indicators and Liver Histomorphological Changes *Animals* 10 (7): 1-18
- Olabanji, R. O., Farinu, G. O., Akinlade, J. A. and Ojebiyi, O. O. 2007.** Growth performance and haematological characteristics of weaner rabbits fed cyanide in processed cassava peel meals on haematological and biochemical indices of growing rabbits. Proc. of the 35th Annual Conference of the Nigeria Society for Animal Production, 212.
- Oyawoye, E. O. and Ogunkunle, M. 1998.** Physiological and biochemical effects of raw Jack beans on broilers. *Proceeding of 23rd Nigeria Society Animal Production* 23: 141-142.

Soltan, M.A. 2009. Growth performance, immune response and carcass traits of broiler chicks fed

on graded levels of palm kernel cake without or with enzyme supplementation. *Livestock Research for Rural Development* 21(3): 29 - 37

Sonaiya, E. B. 2004 Direct assessment of nutrient resources in free range and scavenging systems. *World's Poultry Science Journal* 60:523-535.

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Sunmonu, T.O. and Oloyede, O.B. 2010. Performance and haematological indices in rats exposed to monocrotophos contamination. *Human and Experimental Toxicology* 29(10): 845-850

Togun, V. A. and Oseni, B. S. A. 2005. Effect of low Frequency of administration inclusion

of biscuit dust in broiler finisher diet apparently healthy red Sokoto goats. Proc. of the 27th Annual Conference of Nigeria Society for Animal Production: 50-53.

Tomori, O.A., Saba, A.B. and Dada-Adegbola, H.O. 2007. Antibacterial activity of ethanolic extract of whole fruit of *Lagenaria breviflora Roberts*. *Journal Animal Veterinary Advances* 6: 752-757.

Wang, R., Li D., and Bourne, S., 1998. Can 2000 years of herbal medicine history help us solve problems in the year 2000? Biotechnology in the feed industry: Proceedings of Alltech's 14th Annual Symposium, Kentucky, USA. 273-291.

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